

Corporate Medical Policy

Genetic Testing for Hereditary Pancreatitis AHS – M2079

File Name: genetic_testing_for_hereditary_pancreatitis
Origination: 01/2019
Last CAP Review: 07/2021
Next CAP Review: 07/2022
Last Review: 07/2021

Description of Procedure or Service

Pancreatitis is defined as inflammation of the pancreas that progresses from acute (AP) (sudden onset; duration <6 months) to recurrent acute (RAP) (>1 episode of acute pancreatitis) to chronic (CP) (duration >6 months) (LaRusch, Solomon, & Whitcomb, 2014). This recurrent inflammation can lead to total destruction of the pancreas with subsequent pancreatic insufficiency, secondary diabetes, increased risk for pancreatic cancer and severe unremitting pain (Ravi Kanth & Nageshwar Reddy, 2014).

Hereditary pancreatitis is the early onset form of chronic pancreatitis that is carried in an autosomal dominant pattern with variable penetrance (LaRusch, Barmada, Solomon, & Whitcomb, 2012).

Related Policies

Pancreatic Enzyme Testing for Acute Pancreatitis AHS-G2153
General Genetic Testing, Germline Disorders AHS-M2145
General Genetic Testing, Somatic Disorders AHS-M2146

******Note: This Medical Policy is complex and technical. For questions concerning the technical language and/or specific clinical indications for its use, please consult your physician.***

Policy

BCBSNC will provide coverage for genetic testing for hereditary pancreatitis when it is determined the medical criteria or reimbursement guidelines below are met.

Benefits Application

This medical policy relates only to the services or supplies described herein. Please refer to the Member's Benefit Booklet for availability of benefits. Member's benefits may vary according to benefit design; therefore, member benefit language should be reviewed before applying the terms of this medical policy.

When Genetic Testing for Hereditary Pancreatitis is covered

Genetic testing for hereditary pancreatitis (Note 1) is considered medically necessary in symptomatic patients <20 years old and the individual is presenting with one of the following situations:

- a. Recurrent (two separate, documented episodes with hyperlipasemia) attacks of acute pancreatitis for which there is no identifiable cause
- b. Unexplained chronic pancreatitis

Genetic Testing for Hereditary Pancreatitis AHS – M2079

- c. A family history of recurrent acute pancreatitis, idiopathic chronic pancreatitis, or childhood pancreatitis without a known cause in a first- or second-degree relative
- d. Unexplained episode of pancreatitis in a child that required hospitalization

Note 1: For 5 or more gene tests being run on the same platform, such as multi-gene panel next generation sequencing, please refer to the Laboratory Procedures Medical Policy AHS – R2162

When Genetic Testing for Hereditary Pancreatitis is not covered

Genetic testing for hereditary pancreatitis is considered investigational in all other situations.

Policy Guidelines

Background

Pancreatitis is caused by unregulated trypsin activity within the pancreatic acinar cell or pancreatic duct that leads to pancreatic autodigestion and pancreatic inflammation (Lerch & Gorelick, 2000; Whitcomb, 1999). Under acinar cell stress (e.g., hyperstimulation, intracellular hypercalcemia), intracellular trypsinogen is likely converted to trypsin, which activates other digestive enzymes causing injury. Injury releases immune system-activating molecules that cause an initial acute inflammatory response, followed by recruitment of tissue macrophages and activated pancreatic stellate cells. Recurrent injury leads to chronic pancreatitis and fibrosis, mediated by pancreatic stellate cells (Werlin et al., 2015).

Chronic pancreatitis (CP) is a progressive inflammatory disease in which the pancreatic tissue is destroyed over time and replaced by fibrous tissue. The process of fibrosis usually leads to progressive worsening in the structural integrity of the pancreas, changes in arrangement, and composition of the islets, and deformation of the large ducts, eventually resulting in the impairment of both exocrine and endocrine functions (Brock, Nielsen, Lelic, & Drewes, 2013). The annual incidence of the disease has been estimated to be 5-10 per 100,000 persons (Molven, Njolstad, & Weiss, 2015). The main symptom of CP is pain; however, it is highly variable in character, frequency, and severity (Mullady et al., 2011; Whitcomb et al., 2008). Therapeutic efforts are mostly aimed at extracting stones and decompressing pancreatic ducts to achieve ideal drainage of the pancreatic duct (Li et al., 2010; Tandan & Nageshwar Reddy, 2013).

The etiologies of chronic pancreatitis are classified by the TIGAR-O system into alcoholism, hyperlipidemia, obstructive damage caused by trauma or congenital anomalies, hereditary pancreatitis, autoimmune pancreatitis, and idiopathic (Etemad & Whitcomb, 2001; Sun et al., 2015). The genetic factors listed in TIGAR-O are *PRSSI* (listed as “cationic trypsinogen”), *CFTR*, *SPINK1*, and alpha-1-antitrypsin (listed as “possible”) (Etemad & Whitcomb, 2001). TIGAR-O Version 2 was published in 2019, and lists *PRSSI*, *CFTR*, *SPINK1*, *CTRC*, *CASR*, and *CEL* as genetic factors, as well as some modifier genes such as *CLDN2* (D. C. Whitcomb, 2019).

Hereditary pancreatitis (HP) presents as an autosomal dominant chronic pancreatitis with variable penetrance. This variability has been attributed to a genetic predisposition to chronic pancreatitis with the additive effects of environmental and inherited factors. Most genes associated with HP either directly encode components of the trypsin system of the exocrine pancreas or are likely to perturb this system indirectly. The phenotype of HP is increased susceptibility to acute pancreatitis, resulting in chronic pancreatitis (including pancreatic fibrosis, chronic pain, maldigestion, and diabetes mellitus) occurring in at least 50%. The risk of pancreatic cancer is also increased (D.C. Whitcomb, 2019).

Genes Linked to Hereditary Pancreatitis

PRSSI encodes trypsin-1 (cationic trypsinogen), a major pancreatic digestive enzyme. Mutations in *PRSSI* typically result in a trypsin protein that is either prematurely activated or resistant to

Genetic Testing for Hereditary Pancreatitis AHS – M2079

degradation (LaRusch et al., 2012; Masson, Le Marechal, Delcenserie, Chen, & Ferec, 2008), causing autosomal dominant pancreatitis in 60%-100% of families with hereditary pancreatitis (LaRusch & Whitcomb, 2011). “The age of onset for *PRSS-1* related HP ranges from 10 to 12 years” (Hasan, Moscoso, & Kastrinos, 2018).

SPINK1 encodes serine protease inhibitor, Kazel-type 1, a trypsin inhibitor that is upregulated by inflammation (Grendell, 2003). It is not a typical susceptibility gene for acute pancreatitis, but rather a susceptibility gene for the chronic pancreatitis that follows acute pancreatitis.

CTRC encodes chymotrypsin C. Prematurely activated trypsin is destroyed by *CTRC* by acting on the molecule within the calcium-binding loop in the absence of calcium and, therefore, is a crucial candidate gene in the pathogenesis of pancreatitis (Szmola & Sahin-Toth, 2007).

CASR encodes calcium sensing receptor, mutations of which can cause increased calcium ion levels increasing trypsin activation and failed trypsin degradation (Whitcomb, 2004).

CFTR encodes the cystic fibrosis transmembrane conductance protein. Mutations are associated with recurrent acute and chronic pancreatitis since dysfunctional *CFTR* can result in retention of zymogens that can become active and result in pancreatitis (LaRusch & Whitcomb, 2011).

CLDN2 encodes claudin-2, a tight-junction protein that seals the space between epithelial cells. Normally expressed in the proximal pancreatic duct, *CLDN2* is thought to facilitate the transport of water and sodium into the duct to match the chloride and bicarbonate that are actively secreted by pancreatic duct cells through *CFTR*. It is strongly associated with alcohol-related chronic pancreatitis rather than recurrent acute pancreatitis (Ravi Kanth & Nageshwar Reddy, 2014).

CPA1 encodes carboxypeptidase A1; mutated *CPA1* is associated with nonalcoholic chronic pancreatitis, especially with an early age of onset (Witt et al., 2013). Risk for chronic pancreatitis unrelated to trypsin activation appears to be related to endoplasmic reticulum stress from pathogenic *CPA1* variants that alter protein folding, triggering the unfolded protein response.

MYO9B gene and the two tight-junction adaptor genes, ***PARD3*** and ***MAGI2***, have been linked to gastrointestinal permeability. Impairment of the mucosal barrier plays an important role in the pathophysiology of acute pancreatitis (Nijmeijer et al., 2013).

CEL encodes carboxyl-ester lipase, and *CEL* mutations can cause an autosomal dominant syndrome of maturity-onset diabetes of the young (MODY) and exocrine pancreatic dysfunction (Molven et al., 2015).

Syndromes that Include Pancreatitis or Pancreatic Insufficiency

Several genes are associated with rare disorders in which pancreatitis or pancreatic insufficiency is part of their phenotype (Durie, 1996; Lerch, Zenker, Turi, & Mayerle, 2006).

Genetic Testing for Hereditary Pancreatitis AHS – M2079

Disorders	Genetic Causes	Consequence(s)	Source Citation
Shwachman-Diamond syndrome	SBDS, DNAJC21, EFL1, and SRP54	affect RNA function	(Nelson & Myers, 2008)
Mitochondrial(mt)DNA deletion syndromes, including Kearns-Sayre syndrome (KSS), Pearson syndrome, and progressive external ophthalmoplegia (PEO)	Multiple possible mitochondrial genetic etiologies, including SLC25A4, TWNK, POLG, TYMP, OPA1, RRM2B, DNA2, and MT-TL1	defective oxidative phosphorylation	(Goldstein & Falk 2003)
Carboxyl ester lipase (CEL-MODY)	CEL	pancreatic exocrine, endocrine dysfunction, and chronic pancreatitis	(O'Neill, Stumpf, & McKusick, 2013)
Johanson-Blizzard syndrome	UBD1	protein synthesis	(Kniffin & McKusick, 2012)

As the number of genes and mutations involved in the onset and progression of pancreatitis becomes higher (Ooi, Gonska, Durie, & Freedman, 2010; Walker, Warren, Gawn, & Jiao, 2013), the time and cost of screening and sequencing specific genes continues to increase. However, massive parallel sequencing or next generation sequencing (NGS) is becoming standardized (Ballard et al., 2015), and the cost per patient is rapidly dropping (Palermo et al., 2016). NGS includes whole genome sequencing, whole exome sequencing (WES) and other methods. Because the cost of WES is now less than the cost of sequencing *CFTR*, use of this technology is becoming an attractive alternative to classic targeted gene sequencing or mutation specific genotyping for a genetic counseling workup (LaRusch et al., 2012). In response to this accelerating development of sequencing techniques, several firms have created genetic panels focusing on hereditary pancreatitis. For example, Invitae offers a six-gene panel (*CASR*, *CFTR*, *CPA1*, *CTRC*, *PRSS1*, *SPINK1*) for chronic pancreatitis (Invitae). Other firms offering proprietary panels include ARUP Laboratories (4 genes), LabCorp (3 genes), and Ambry (6 genes) (Ambry, 2020; ARUP, 2020; LabCorp, 2020). Still other firms evaluate as many as 12 genes and more (D.C. Whitcomb, 2019).

Clinical Validity and Utility

Testing for mutations in the *PRSS1*, *SPINK*, and *CFTR* genes is usually done by either direct sequence analysis or next generation sequencing, both of which have high analytic validity. Several studies have evaluated the clinical validity of genetic testing (Applebaum-Shapiro et al., 2001; Ceppa et al., 2013; Poddar, Yachha, Mathias, & Choudhuri, 2015; Sultan, Werlin, & Venkatasubramani, 2012). One limitation with some studies was lack of inclusion of patients with clinically defined hereditary pancreatitis. Hence, the true clinical sensitivity and specificity of genetic testing in hereditary pancreatitis cannot be accurately determined and needs to be further researched. Similarly, there is a lack of published literature on the clinical utility of testing. Further research is required to evaluate how genetic testing will impact patient management decision and clinical outcomes.

Genetic Testing for Hereditary Pancreatitis AHS – M2079

Kumar et al (2016) sought to characterize and identify risk factors associated with acute recurrent pancreatitis (ARP) and CP in childhood in a multinational cross-sectional study (INSPPIRE). The authors analyzed 301 children with ARP or CP. They found that “At least 1 gene mutation in pancreatitis-related genes was found in 48% of patients with ARP vs 73% of patients with CP. Children with PRSS1 or SPINK1 mutations were more likely to present with CP compared with ARP (PRSS1: OR = 4.20 and SPINK1: OR = 2.30). Obstructive risk factors presented in 33% in both groups, but toxic/metabolic risk factors were more common in children with ARP (21% overall; 26% ARP, 15% CP). They concluded that “The high disease burden in pediatric CP underscores the importance of identifying predisposing factors for progression of ARP to CP in children (Kumar et al., 2016).”

Gabarczyk (2017) et al also found that CTRC variants are strong CP risk factors in pediatric patients. The authors investigated 136 pediatric patients with CP and compared them to 401 controls. They showed that p.Arg254Trp (4.6%) and p.Lys247_Arg254del (5.3%) heterozygous mutations are frequent and significantly associated with CP risk in pediatric patients (odds ratio [OR]=19.1; 95% CI 2.8-160; P=0.001 and OR=5.5; 95% CI 1.6-19.4; P=0.001, respectively). The c.180TT genotype of common p.Gly60Gly variant was found to be a strong and independent CP risk factor (OR=23; 95% CI 7.7-70; P<0.001) with effect size comparable to p.Arg254Trp mutation (Grabarczyk et al., 2017).

Schwarzenberg et al evaluated the genetic spectrum of CP. 76 CP patients were examined, and 51 were found to have a genetic risk factor for CP. Of these 51 mutations, 33 were a PRSS1 mutation, 14 were a SPINK1 mutation, 11 were a CFTR mutation, and 2 were a CTRC mutation. The final 25 patients were found to have an obstructive risk factor (Schwarzenberg et al., 2015).

Zou et al evaluated the prevalence of four CP-related genes (*SPINK1*, *PRSS1*, *CTRC*, *CFTR*) in Han Chinese patients. The authors performed next-generation sequencing on 1061 patients and 1196 controls. The 1061 patients were further divided into three categories, idiopathic CP (ICP, 715 patients), alcoholic CP (ACP, 206), and smoking-associated CP (SCP, 140). The impact of rare pathogenic variants on age of onset and clinical outcomes was evaluated. Rare pathogenic variants were found in 535 CP patients compared to 71 controls. Mutation positive patients were found to have earlier age of onset as well additional clinical features such as pancreatic stones and diabetes mellitus compared to mutation negative ICP patients. Overall, pathogenic variants were found in 57.1% of ICP patients, compared to 39.8% of ACP patients and 32.1% of SCP patients. The authors concluded that rare pathogenic variants “significantly” influenced age of onset and clinical outcomes of CP (Zou et al., 2018).

Nabi et al. (2020) evaluated 239 children in a prospective study from January 2015 to May 2018 to examine genetic risk factors in children with idiopathic acute recurrent pancreatitis (IARP). Among the enrollees, 85.35% children had IARP, and found that family history of pancreatitis was found among 4.6% of participants. For specific genes, “mutations/polymorphisms in at least 1 gene were identified in 89.5% (129/144) children including *SPINK1* in 41.9%, *PRSS1* (rs10273639) in 58.2%, *CTRC* in 25.6%, *CTSB* in 54.9%, *CLDN2* in 72.9%, and *CFTR* in 2.3%.” This conveys the overlapping genetic nature of IARP with related genes in HP, making genetic testing important for managing potential disease progression.

Suzuki, Minowa, Nakano, Isayama, and Shimizu (2020) investigated the currently understood genetic abnormalities in pancreatitis, and found that “patients with these genetic predispositions [*PRSS1* and *SPINK1* genes], both children and adults, have often been initially diagnosed with idiopathic acute pancreatitis, in approximately 20-50% pediatric cases and 28-0% of adult cases... Patients with chronic pancreatitis (CP) due to *SPINK1* gene mutation and HP patients have a potentially high risk of pancreatic exocrine insufficiency, diabetes mellitus, and of particular importance, pancreatic cancer.” This conveys the continuously emphasized clinical utility of genetic testing to pursue opportunities for counselling and symptom management with disease

Genetic Testing for Hereditary Pancreatitis AHS – M2079

progression, despite not having gene therapy options for directly targeting HP causing and associated genes (Suzuki et al., 2020).

Weiss, Laemmerhirt, and Lerch (2020) discussed the potential pitfalls from using next generation sequencing (NGS) to diagnose *PRSS1* mutations in chronic pancreatitis. Due to the “high degree of DNA sequence homology (>91%) between *PRSS1* and other members of the trypsinogen multigene family,” there may be erroneous diagnoses of pathologic chronic pancreatitis among patients with benign variants of other *PRSS1*- related genes, like *PRSS2* or *PRSS3P2*. The researchers concluded that sequence homology “can confound the mapping of short NGS reads to a reference genome and lead to technical artefacts.” They recommend “careful clinical evaluation, pretest and post-test genetic counselling and confirmation of NGS test results by Sanger sequencing” to confirm a diagnosis of genetically mutated chronic pancreatitis. This presented the precautions that must be accounted for when utilizing genetic testing for hereditary pancreatitis (Weiss et al., 2020).

Guidelines and Recommendations

A Consensus Committees of the **European Registry of Hereditary Pancreatic Diseases** the **Midwest Multi-Center Pancreatic Study Group**, and the **International Association of Pancreatology**, developed guidelines for genetic testing of the *PRSS1* gene and genetic counseling for HP (Ellis, Lerch, & Whitcomb, 2001). The recommended indications for symptomatic patients included:

- Recurrent (two separate, documented episodes with hyperlipasemia) attacks of acute pancreatitis for which there is no explanation (anatomical anomalies, ampullary or main pancreatic strictures, trauma, viral infection, gallstones, alcohol, drugs, hyperlipidaemia, etc.)
- Unexplained chronic pancreatitis
- A family history of pancreatitis in a first- or second-degree relative
- Unexplained episode of pancreatitis in a child that required hospitalization

Predictive (presymptomatic) genetic testing of unaffected relatives is considered more complex. Predictive testing is recommended only for individuals with a first-degree relative with a defined HP gene mutation, and who are over 16 years of age and capable of making an independent a fully informed decision (Ellis et al., 2001).

American Society of Clinical Oncology (ASCO)

ASCO states that “genetic testing is sometimes considered for patients who develop recurrent pancreatitis at young ages. Genetic testing is available for mutations in the *PRSS1*, *SPINK1*, and *CFTR* genes (ASCO, 2020).”

American College of Gastroenterology (ACG) (2013, 2020)

In 2013, the ACG issued guidelines for the management of acute pancreatitis. They include the following recommendation: “genetic testing may be considered in young patients (< 30 years old) if no cause is evident and a family history of pancreatic disease is present (conditional recommendation, low quality of evidence)”. ACG also states that “the role of genetic testing in AP has yet to be determined, but may be useful in patients with more than one family member with pancreatic disease. Individuals with IAP and a family history of pancreatic diseases should be referred for formal genetic counseling” (Tenner, Baillie, DeWitt, & Vege, 2013).

In 2020, the ACG published an update on chronic pancreatitis. In it, they recommend genetic testing in patients “with clinical evidence of a pancreatitis-associated disorder or possible CP [chronic pancreatitis] in which the etiology is unclear, especially in younger patients (strong

Genetic Testing for Hereditary Pancreatitis AHS – M2079

recommendation, low quality of evidence)”. The guideline goes on to state that “at minimum, patients with idiopathic CP should be evaluated for *PRSS1*, *SPINK1*, *CFTR*, and *CTRC* gene mutation analysis...” The guideline mentions that assessment of germline mutations is primarily for prognostic and therapeutic purposes, rather than diagnostic (Gardner et al., 2020).

United European Gastroenterology (2017)

The United European Gastroenterology published evidence-based guidelines (Lohr et al., 2017) for the diagnosis and therapy of chronic pancreatitis which recommend:

“All patients with a family history or early onset disease (<20 years) should be offered genetic testing for associated variants.”

“Genetic screening for every CP patient cannot be recommended since alcohol abuse is the predominant cause of the disease in up to 60% of adult cases.”

“In patients with early onset CP, genetic screening can be offered after informed consent.”

“In patients with alcoholic CP, routine genetic testing cannot be recommended.”

The working group also noted that “variants in *SPINK1* and *CTRC*, and to a lesser extent, common single-nucleotide polymorphisms (SNPs) in the *PRSS1* and *CLDN2-MORC4* loci, are associated with alcoholic CP” (Lohr et al., 2017).

European Pancreatic Club/ Hungarian Pancreatic Study Group (EPC/HPSG, 2018)

The European Pancreatic Club, in collaboration with the Hungarian Pancreatic Study Group organized a consensus guideline meeting on the diagnosis and management of pancreatitis in the pediatric population which state (Parniczky et al., 2018):

“Pediatric AP and RAP often develop in the background of genetic susceptibility and genetic testing is warranted in patients with a second episode of idiopathic AP or first episode of idiopathic AP and a family history of AP or CP. Full sequence analysis of *PRSS1*, *SPINK1*, *CTRC*, *CPA1* and *CFTR* gene exons and exon-intron boundaries and testing for the pathogenic *CEL* hybrid allele are recommended.”

“Variants in the *PRSS1* and *CPA1* genes may be associated with a family history of pancreatitis or even autosomal dominant hereditary pancreatitis. Children with a single episode of AP are at risk for developing a second episode. However, genetic testing is cumbersome and expensive. There is usually no therapeutic consequence, but it may assist in long term prognosis.”

“The presence of mutations in the above mentioned genes increases the risk of ARP and CP. Hereditary pancreatitis associated with mutations in *PRSS1*, especially p.R122H that could considerably increase the risk of pancreatic adenocarcinoma. Knowing the genetic risk factors may not alter the therapy, but it helps to understand the disease's etiological background for the disease and may lead to future targeted investigation (Parniczky et al., 2018).”

International Study Group of Pediatric Pancreatitis: In search for a cuRE (INSPPIRE) Consortium (2017)

This group was formed “to collect detailed information on a cohort of children with ARP and CP with the aim to fill gaps in knowledge and improve clinical care”. Their genetic testing-related guidelines are listed below:

Genetic Testing for Hereditary Pancreatitis AHS – M2079

- “The search for a genetic cause of ARP or CP should include a sweat chloride test (even if newborn screening for cystic fibrosis (CF) is negative) and PRSS1 gene mutation testing. Genetic testing for CF should be considered if a sweat test is unable to be performed.”
- “Mutation analysis of the genes SPINK1, CFTR and CTRC may identify risk factors for ARP or CP.”
- “Patients with ARP or CP and a sweat test ≤ 60 mmol/L should have expanded CFTR mutation testing done if there is no other identified cause of their pancreatic disease (such as a PRSS1 mutation or a clear obstructive etiology) (Garipey et al., 2017).”

National Comprehensive Cancer Network (NCCN)

The NCCN lists chronic pancreatitis as a risk factor for pancreatic adenocarcinoma and specifically lists *PRSS1*, *SPINK1*, and *CFTR* as contributing genes to familial pancreatitis (NCCN, 2021).

International Association of Pancreatology, American Pancreatic Association, Japan Pancreas Society, and European Pancreatic Club (Greenhalf et al., 2020)

In 2020, the International Association of Pancreatology, the American Pancreatic Association, the Japan Pancreas Society, and European Pancreatic Club released a set of international consensus guidelines on surveillance for pancreatic cancer in the setting of chronic pancreatitis. Though the working group did not explicitly endorse or oppose genetic testing, it was clear that due to the recommendations separated by genetic variants within chronic pancreatitis, genetic testing would become critical for surveillance. With regards to the conditions by which hereditary pancreatitis would warrant surveillance for cancer, the working group stated:

- “The risk of pancreatic cancer in affected individuals with an autosomal dominant history of hereditary pancreatitis due to inherited *PRSS1* mutations is high enough to justify surveillance. *Quality assessment: high; recommendation: strong*”
- “The risk of pancreatic cancer in affected individuals with an autosomal dominant history of hereditary pancreatitis but without *PRSS1* mutations is high enough to justify surveillance. *Quality assessment: moderate; recommendation: weak*”
- “The risk of pancreatic cancer in patients with chronic pancreatitis associated with *SPINK1* p. N34S is not high enough to justify screening or surveillance. *Quality assessment: moderate; recommendation: strong*”
- “The risk of pancreatic cancer in patients with chronic pancreatitis associated with other germline mutations including those of *CFTR*, *CTRC*, *CPA1*, and *CEL*, is not high enough to justify screening or surveillance. *Quality assessment: moderate; recommendation: conditional*” (Greenhalf et al., 2020).

National Institute for Health and Care Excellence (NICE)

NICE updated their guidelines on pancreatitis in December 2020. With regards to genetic testing for hereditary pancreatitis (acute) and patient information, NICE stated the following:

Genetic Testing for Hereditary Pancreatitis AHS – M2079

“Give people with pancreatitis, and their family members or carers (as appropriate), written and verbal information on the following, where relevant, as soon as possible after diagnosis:

- pancreatitis and any proposed investigations and procedures, using diagrams
- hereditary pancreatitis, and pancreatitis in children, including specific information on genetic counselling, genetic testing, risk to other family members, and advice on the impact of their pancreatitis on life insurance and travel
- the long-term effects of pancreatitis, including effects on the person's quality of life the harm caused to the pancreas by smoking or alcohol.”

For an individual with chronic pancreatitis, NICE recognizes that the cause may not be alcohol-related, but can include “genetic factors; autoimmune disease, in particular IgG4 disease; metabolic causes; [and] structural or anatomical factors” (NICE, 2020).

Applicable Federal Regulations

A search on the FDA website on April 22, 2021 for “pancreatitis” yielded no genetic results. Additionally, many labs have developed specific tests that they must validate and perform in house. These laboratory-developed tests (LDTs) are regulated by the Centers for Medicare and Medicaid (CMS) as high-complexity tests under the Clinical Laboratory Improvement Amendments of 1988 (CLIA '88). As an LDT, the U. S. Food and Drug Administration has not approved or cleared this test; however, FDA clearance or approval is not currently required for clinical use.

Billing/Coding/Physician Documentation Information

This policy may apply to the following codes. Inclusion of a code in this section does not guarantee that it will be reimbursed. For further information on reimbursement guidelines, please see Administrative Policies on the Blue Cross Blue Shield of North Carolina web site at www.bcbsnc.com. They are listed in the Category Search on the Medical Policy search page.

Applicable service codes: 81222, 81223, 81224, 81401, 81404, 81405, 81479

BCBSNC may request medical records for determination of medical necessity. When medical records are requested, letters of support and/or explanation are often useful, but are not sufficient documentation unless all specific information needed to make a medical necessity determination is included.

Scientific Background and Reference Sources

- Ambry. (2020). Pancreatitis panel. Retrieved from <https://www.ambrygen.com/clinician/genetic-testing/69/exome-and-general-genetics/pancreatitis-panel>
- Applebaum-Shapiro, S. E., Finch, R., Pfutzer, R. H., Hepp, L. A., Gates, L., Amann, S., . . . Whitcomb, D. C. (2001). Hereditary pancreatitis in North America: the Pittsburgh-Midwest Multi-Center Pancreatic Study Group Study. *Pancreatology*, 1(5), 439-443. Retrieved from <http://dx.doi.org/>
- ARUP. (2020, December 7). Pancreatitis Panel (CFTR, CTRC, PRSS1, SPINK1) Sequencing. Retrieved from <https://ltd.aruplab.com/Tests/Pub/2010876ter>
- ASCO. (2020). Hereditary Pancreatitis. Retrieved from <https://www.cancer.net/cancer-types/hereditary-pancreatitis>
- Ballard, D. D., Flueckiger, J. R., Fogel, E. L., McHenry, L., Lehman, G. A., Watkins, J. L., . . . Cote, G. A. (2015). Evaluating Adults With Idiopathic Pancreatitis for Genetic Predisposition: Higher Prevalence of Abnormal Results With Use of Complete Gene Sequencing. *Pancreas*, 44(1), 116-121. doi:10.1097/mpa.0000000000000225
- Brock, C., Nielsen, L. M., Lelic, D., & Drewes, A. M. (2013). Pathophysiology of chronic pancreatitis. *World J Gastroenterol*, 19(42), 7231-7240. doi:10.3748/wjg.v19.i42.7231

Genetic Testing for Hereditary Pancreatitis AHS – M2079

- Ceppa, E. P., Pitt, H. A., Hunter, J. L., Leys, C. M., Zyromski, N. J., Rescorla, F. J., . . . Lehman, G. A. (2013). Hereditary pancreatitis: endoscopic and surgical management. *J Gastrointest Surg*, *17*(5), 847-856; discussion 856-847. doi:10.1007/s11605-013-2167-8
- Durie, P. R. (1996). Inherited and congenital disorders of the exocrine pancreas. *Gastroenterologist*, *4*(3), 169-187. Retrieved from <http://dx.doi.org/>
- Ellis, I., Lerch, M. M., & Whitcomb, D. C. (2001). Genetic testing for hereditary pancreatitis: guidelines for indications, counselling, consent and privacy issues. *Pancreatology*, *1*(5), 405-415. Retrieved from <https://www.karger.com/Article/PDF/55840>
- Etemad, B., & Whitcomb, D. C. (2001). Chronic pancreatitis: diagnosis, classification, and new genetic developments. *Gastroenterology*, *120*(3), 682-707. Retrieved from <http://dx.doi.org/>
- Gardner, T. B., Adler, D. G., Forsmark, C. E., Sauer, B. G., Taylor, J. R., & Whitcomb, D. C. (2020). ACG Clinical Guideline: Chronic Pancreatitis. *American Journal of Gastroenterology*, *115*(3). Retrieved from https://journals.lww.com/ajg/Fulltext/2020/03000/ACG_Clinical_Guideline_Chronic_Pancreatitis.9.aspx
- Garipey, C. E., Heyman, M. B., Lowe, M. E., Pohl, J. F., Werlin, S. L., Wilschanski, M., . . . Uc, A. (2017). Causal Evaluation of Acute Recurrent and Chronic Pancreatitis in Children: Consensus From the INSPPIRE Group. *J Pediatr Gastroenterol Nutr*, *64*(1), 95-103. doi:10.1097/mpg.0000000000001446
- Goldstein, A., & Falk, M. (2003, 01/31/2019). Mitochondrial DNA Deletion Syndromes. *GeneReviews*(r). Retrieved from <https://www.ncbi.nlm.nih.gov/books/NBK1203/>
- Grabarczyk, A. M., Oracz, G., Wertheim-Tysarowska, K., Kujko, A. A., Wejnarska, K., Kolodziejczyk, E., . . . Rygiel, A. M. (2017). Chymotrypsinogen C Genetic Variants, Including c.180TT, Are Strongly Associated With Chronic Pancreatitis in Pediatric Patients. *J Pediatr Gastroenterol Nutr*, *65*(6), 652-657. doi:10.1097/mpg.0000000000001767
- Greenhalf, W., Lévy, P., Gress, T., Rebours, V., Brand, R. E., Pandol, S., . . . Neoptolemos, J. (2020). International consensus guidelines on surveillance for pancreatic cancer in chronic pancreatitis. Recommendations from the working group for the international consensus guidelines for chronic pancreatitis in collaboration with the International Association of Pancreatology, the American Pancreatic Association, the Japan Pancreas Society, and European Pancreatic Club. *Pancreatology*, *20*(5), 910-918. doi:10.1016/j.pan.2020.05.01
- Grendell, J. H. (2003). Genetic factors in pancreatitis. *Curr Gastroenterol Rep*, *5*(2), 105-109. Retrieved from <http://dx.doi.org/>
- [Hasan, A., Moscoso, D. I., & Kastrinos, F. \(2018\). The Role of Genetics in Pancreatitis. *Gastrointest Endosc Clin N Am*, *28*\(4\), 587-603. doi:10.1016/j.giec.2018.06.001](#)
- Invitae. Invitae Chronic Pancreatitis Panel. Retrieved from https://www.invitae.com/en/physician/tests/01745/#info-panel-disorders_tested
- Kniffin, C., & McKusick, V. (2012, 09/15/2016). JOHANSON-BLIZZARD SYNDROME; JBS. *Online Mendelian Inheritance in Man*. Retrieved from <https://omim.org/entry/243800>
- Kumar, S., Ooi, C. Y., Werlin, S., Abu-El-Haija, M., Barth, B., Bellin, M. D., . . . Uc, A. (2016). Risk Factors Associated With Pediatric Acute Recurrent and Chronic Pancreatitis: Lessons From INSPPIRE. *JAMA Pediatr*, *170*(6), 562-569. doi:10.1001/jamapediatrics.2015.4955
- LabCorp. (2020). Pancreatitis: Three-gene Profile (PRSS1, SPINK1, CFTR) (Full Gene Sequencing). Retrieved from <https://www.labcorp.com/tests/252794/pancreatitis-three-gene-profile-i-prss1-spink1-cftr-i-full-gene-sequencing>
- LaRusch, J., Barmada, M. M., Solomon, S., & Whitcomb, D. C. (2012). Whole exome sequencing identifies multiple, complex etiologies in an idiopathic hereditary pancreatitis kindred. *Jop*, *13*(3), 258-262. Retrieved from <http://dx.doi.org/>
- LaRusch, J., Solomon, S., & Whitcomb, D. C. (2014). Pancreatitis Overview. *GeneReviews*. doi:<https://www.ncbi.nlm.nih.gov/books/NBK190101/>
- LaRusch, J., & Whitcomb, D. C. (2011). Genetics of pancreatitis. *Curr Opin Gastroenterol*, *27*(5), 467-474. doi:10.1097/MOG.0b013e328349e2f8
- Lerch, M. M., & Gorelick, F. S. (2000). Early trypsinogen activation in acute pancreatitis. *Med Clin North Am*, *84*(3), 549-563, viii. Retrieved from <https://www.sciencedirect.com/science/article/abs/pii/S002571250570239X>

Genetic Testing for Hereditary Pancreatitis AHS – M2079

- Lerch, M. M., Zenker, M., Turi, S., & Mayerle, J. (2006). Developmental and metabolic disorders of the pancreas. *Endocrinol Metab Clin North Am*, 35(2), 219-241, vii. doi:10.1016/j.ecl.2006.02.004
- Li, Z. S., Wang, W., Liao, Z., Zou, D. W., Jin, Z. D., Chen, J., . . . Wang, L. (2010). A long-term follow-up study on endoscopic management of children and adolescents with chronic pancreatitis. *Am J Gastroenterol*, 105(8), 1884-1892. doi:10.1038/ajg.2010.85
- Lohr, J. M., Dominguez-Munoz, E., Rosendahl, J., Besselink, M., Mayerle, J., Lerch, M. M., . . . Bruno, M. (2017). United European Gastroenterology evidence-based guidelines for the diagnosis and therapy of chronic pancreatitis (HaPanEU). *United European Gastroenterol J*, 5(2), 153-199. doi:10.1177/2050640616684695
- Masson, E., Le Marechal, C., Delcenserie, R., Chen, J. M., & Ferec, C. (2008). Hereditary pancreatitis caused by a double gain-of-function trypsinogen mutation. *Hum Genet*, 123(5), 521-529. doi:10.1007/s00439-008-0508-6
- Molven, A., Njolstad, P. R., & Weiss, F. U. (2015). Lipase gene fusion: a new route to chronic pancreatitis. In *Oncotarget* (Vol. 6, pp. 30443-30444). United States.
- Mullady, D. K., Yadav, D., Amann, S. T., O'Connell, M. R., Barmada, M. M., Elta, G. H., . . . Anderson, M. A. (2011). Type of pain, pain-associated complications, quality of life, disability and resource utilisation in chronic pancreatitis: a prospective cohort study. *Gut*, 60(1), 77-84. doi:10.1136/gut.2010.213835
- Nabi, Z., Talukdar, R., Venkata, R., Aslam, M., Shava, U., & Reddy, D. N. (2020). Genetic Evaluation of Children with Idiopathic Recurrent Acute Pancreatitis. *Dig Dis Sci*, 65(10), 3000-3005. doi:10.1007/s10620-019-06026-2
- NCCN. (2019). Pancreatic Adenocarcinoma. Retrieved from https://www.nccn.org/professionals/physician_gls/pdf/pancreatic.pdf
- Nelson, A., & Myers, K. (2008, 10/18/2018). Shwachman-Diamond Syndrome. *GeneReviews(r)*. Retrieved from [https://www.ncbi.nlm.nih.gov/books/NBK1756/NICE.\(2020,December16\).Pancreatitis.Retrievedfromhttps://www.nice.org.uk/guidance/ng104/chapter/Recommendations](https://www.ncbi.nlm.nih.gov/books/NBK1756/NICE.(2020,December16).Pancreatitis.Retrievedfromhttps://www.nice.org.uk/guidance/ng104/chapter/Recommendations)
- Nijmeijer, R. M., van Santvoort, H. C., Zhernakova, A., Teller, S., Scheiber, J. A., de Kovel, C. G., . . . Wijmenga, C. (2013). Association analysis of genetic variants in the myosin IXB gene in acute pancreatitis. *PLoS One*, 8(12), e85870. doi:10.1371/journal.pone.0085870
- O'Neill, M., Stumpf, A., & McKusick, V. (2013, 09/10/2013). MATURITY-ONSET DIABETES OF THE YOUNG, TYPE 8, WITH EXOCRINE DYSFUNCTION; MODY8. *Online Mendelian Inheritance in Man*. Retrieved from <https://omim.org/entry/609812>
- Ooi, C. Y., Gonska, T., Durie, P. R., & Freedman, S. D. (2010). Genetic testing in pancreatitis. *Gastroenterology*, 138(7), 2202-2206, 2206.e2201. doi:10.1053/j.gastro.2010.04.022
- Palermo, J. J., Lin, T. K., Hornung, L., Valencia, C. A., Mathur, A., Jackson, K., . . . Abu-El-Haija, M. (2016). Genophenotypic Analysis of Pediatric Patients With Acute Recurrent and Chronic Pancreatitis. *Pancreas*, 45(9), 1347-1352. doi:10.1097/mpa.0000000000000655
- Parniczky, A., Abu-El-Haija, M., Husain, S., Lowe, M., Oracz, G., Sahin-Toth, M., . . . Hegyi, P. (2018). EPC/HPSG evidence-based guidelines for the management of pediatric pancreatitis. *Pancreatolgy*, 18(2), 146-160. doi:10.1016/j.pan.2018.01.001
- Poddar, U., Yachha, S. K., Mathias, A., & Choudhuri, G. (2015). Genetic predisposition and its impact on natural history of idiopathic acute and acute recurrent pancreatitis in children. *Dig Liver Dis*, 47(8), 709-714. doi:10.1016/j.dld.2015.04.012
- Ravi Kanth, V., & Nageshwar Reddy, D. (2014). Genetics of acute and chronic pancreatitis: An update. *World J Gastrointest Pathophysiol*, 5(4), 427-437. doi:10.4291/wjgp.v5.i4.427
- Schwarzenberg, S. J., Bellin, M., Husain, S. Z., Ahuja, M., Barth, B., Davis, H., . . . Uc, A. (2015). Pediatric chronic pancreatitis is associated with genetic risk factors and substantial disease burden. *J Pediatr*, 166(4), 890-896.e891. doi:10.1016/j.jpeds.2014.11.019
- Sultan, M., Werlin, S., & Venkatasubramani, N. (2012). Genetic prevalence and characteristics in children with recurrent pancreatitis. *J Pediatr Gastroenterol Nutr*, 54(5), 645-650. doi:10.1097/MPG.0b013e31823f0269
- Sun, C., Liu, M. Y., Liu, X. G., Hu, L. H., Xia, T., Liao, Z., & Li, Z. S. (2015). Serine Protease Inhibitor Kazal Type 1 (SPINK1) c.194+2T > C Mutation May Predict Long-term Outcome of

Genetic Testing for Hereditary Pancreatitis AHS – M2079

- Endoscopic Treatments in Idiopathic Chronic Pancreatitis. *Medicine (Baltimore)*, 94(47), e2046. doi:10.1097/md.0000000000002046
- Suzuki, M., Minowa, K., Nakano, S., Isayama, H., & Shimizu, T. (2020). Genetic Abnormalities in Pancreatitis: An Update on Diagnosis, Clinical Features, and Treatment. *Diagnostics (Basel, Switzerland)*, 11(1), 31. doi:10.3390/diagnostics11010031
- Szmola, R., & Sahin-Toth, M. (2007). Chymotrypsin C (caldecrin) promotes degradation of human cationic trypsin: identity with Rinderknecht's enzyme Y. *Proc Natl Acad Sci U S A*, 104(27), 11227-11232. doi:10.1073/pnas.0703714104
- Tandan, M., & Nageshwar Reddy, D. (2013). Endotherapy in chronic pancreatitis. *World J Gastroenterol*, 19(37), 6156-6164. doi:10.3748/wjg.v19.i37.6156
- Tenner, S., Baillie, J., DeWitt, J., & Vege, S. S. (2013). American College of Gastroenterology guideline: management of acute pancreatitis. *Am J Gastroenterol*, 108(9), 1400-1415; 1416. doi:10.1038/ajg.2013.218
- Walker, N. F., Warren, O. J., Gawn, L., & Jiao, L. R. (2013). The role of genetic testing in management of hereditary chronic pancreatitis. *JRSM Short Rep*, 4(1), 6. doi:10.1258/shorts.2012.012071
- Weiss, F. U., Laemmerhirt, F., & Lerch, M. M. (2020). Next generation sequencing pitfalls in diagnosing trypsinogen (PRSS1) mutations in chronic pancreatitis. *Gut*. doi:10.1136/gutjnl-2020-322864
- Werlin, S., Konikoff, F. M., Halpern, Z., Barkay, O., Yerushalmi, B., Broide, E., . . . Wilschanski, M. (2015). Genetic and electrophysiological characteristics of recurrent acute pancreatitis. *J Pediatr Gastroenterol Nutr*, 60(5), 675-679. doi:10.1097/mpg.0000000000000623
- Whitcomb, D. C. (1999). Early trypsinogen activation in acute pancreatitis. *Gastroenterology*, 116(3), 770-772. Retrieved from [https://www.gastrojournal.org/article/S0016-5085\(99\)70205-2/fulltext](https://www.gastrojournal.org/article/S0016-5085(99)70205-2/fulltext)
- Whitcomb, D. C. (2004). Mechanisms of disease: Advances in understanding the mechanisms leading to chronic pancreatitis. *Nat Clin Pract Gastroenterol Hepatol*, 1(1), 46-52. doi:10.1038/ncpgasthep0025
- Whitcomb, D. C. (2019). Hereditary pancreatitis. Retrieved from https://www.uptodate.com/contents/hereditary-pancreatitis?search=Hereditary%20pancreatitis&source=search_result&selectedTitle=1~24&usage_type=default&display_rank=1#H2
- Whitcomb, D. C. (2019). Pancreatitis: TIGAR-O Version 2 Risk/Etiology Checklist With Topic Reviews, Updates, and Use Primers. *Clin Transl Gastroenterol*, 10(6), e00027. doi:10.14309/ctg.0000000000000027
- Whitcomb, D. C., Yadav, D., Adam, S., Hawes, R. H., Brand, R. E., Anderson, M. A., . . . Barmada, M. M. (2008). Multicenter approach to recurrent acute and chronic pancreatitis in the United States: the North American Pancreatitis Study 2 (NAPS2). *Pancreatolgy*, 8(4-5), 520-531. doi:10.1159/000152001
- Witt, H., Beer, S., Rosendahl, J., Chen, J. M., Chandak, G. R., Masamune, A., . . . Sahin-Toth, M. (2013). Variants in CPA1 are strongly associated with early onset chronic pancreatitis. *Nat Genet*, 45(10), 1216-1220. doi:10.1038/ng.2730
- Zou, W. B., Tang, X. Y., Zhou, D. Z., Qian, Y. Y., Hu, L. H., Yu, F. F., . . . Liao, Z. (2018). SPINK1, PRSS1, CTSC, and CFTR Genotypes Influence Disease Onset and Clinical Outcomes in Chronic Pancreatitis. *Clin Transl Gastroenterol*, 9(11), 204. doi:10.1038/s41424-018-0069-5

Specialty Matched Consultant Advisory Panel review 7/2019

Medical Director review 7/2019

Specialty Matched Consultant Advisory Panel review 7/2020

Medical Director review 7/2020

Specialty Matched Consultant Advisory Panel review 7/2021

Genetic Testing for Hereditary Pancreatitis AHS – M2079

Medical Director review 7/2021

Policy Implementation/Update Information

- 1/1/2019 BCBSNC will provide coverage for genetic testing for hereditary pancreatitis when it is determined to be medically necessary because the medical criteria and guidelines are met. Medical Director review 1/1/2019. Policy noticed 1/1/2019 for effective date 4/1/2019 (jd)
- 8/13/2019 Specialty Matched Consultant Advisory Panel review 7/2019. Medical Director review 7/2019. (jd)
- 9/10/2019 Reviewed by Avalon 2nd Quarter 2019 CAB. Related Policies added to Description section. Minor revision to When Covered section; removed “Unexplained pancreatitis with” from item 1c. Policy guidelines and references updated. Code table removed from Billing/Coding section. Medical Director review 8/2019. (jd)
- 10/29/19 Wording in the Policy, When Covered, and/or Not Covered section(s) changed from Medical Necessity to Reimbursement language, where needed. (hb)
- 7/28/20 Reviewed by Avalon 2nd Quarter 2020 CAB. Note 1 added to When Covered section. Policy guidelines and references updated. Specialty Matched Consultant Advisory Panel review 7/2020. Medical Director review 7/2020. (jd)
- 8/24/21 Reviewed by Avalon 2nd Quarter 2021 CAB. Policy guidelines and references updated. Specialty Matched Consultant Advisory Panel review 7/2021. Medical Director review 7/2021. (jd)

Medical policy is not an authorization, certification, explanation of benefits or a contract. Benefits and eligibility are determined before medical guidelines and payment guidelines are applied. Benefits are determined by the group contract and subscriber certificate that is in effect at the time services are rendered. This document is solely provided for informational purposes only and is based on research of current medical literature and review of common medical practices in the treatment and diagnosis of disease. Medical practices and knowledge are constantly changing and BCBSNC reserves the right to review and revise its medical policies periodically.